

# Effect of Aqueous Leaf Extract of *Annona senegalensis* on Selected Testicular Function Indices of Wistar Rats

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**Abstract:** This study evaluated the effect of *Annona senegalensis* on selected biochemical and testicular function indices in rats. Twenty Wistar rats (125 – 127 g) were randomly assigned into four experimental groups: A, B, C and D, (n = 5/ group). Animals in group A served as control and was administered distilled water, while animals in groups B, C, and D were given 100 mg/kg body weight (bw), 200 mg/kg bw and 300 mg/kg bw of aqueous leaf extract of *Annona senegalensis* (ALEAS) respectively by oral gavage for 21 days. Data revealed a significant decrease ( $P < 0.05$ ) in the testis-body weight ratio and total protein content in the testes of rats administered ALEAS compared to the control. There was also a significant increase ( $P < 0.05$ ) in testicular cholesterol, glycogen, malondialdehyde (MDA) and catalase (CAT) activity. Additionally, histopathological studies revealed a marked degeneration of seminiferous tubules. Overall, data from the present study suggest that aqueous leaf extract of *Annona senegalensis* may possess the potential to adversely affect testicular function in rat.

**Keywords:** *Annona senegalensis*, Testes, Testicular Function, Histopathology, Rat

## 1. Introduction

*Annona senegalensis* is a subtropical plant [1] that has been implicated for the treatment of chest pain, coughs, anaemia, urinary tract infection [2, 3], cancer treatment [4, 5], diarrhoea, dysentery [6, 7], antritis and rheumatism [8, 9]. The plant has also been shown to be effective as an antiulcer/antacid, smooth muscle relaxant [10], antibacterial [3], antiprotozoal [11], molluscicidal [12] and hormonal activities [13]. It can be used as a pain reliever, a stimulant, and antihelminthic [14, 15]. There are unsubstantiated claims that the plant is being used locally to enhance sexual performance and boost fertility in male. The rate of spermatogenesis and the quality of the sperm is necessary for fertility in a man. The testis which is always in pair in human are responsible for the production of semen which contain the sperm cell. Albeit, studies showing the effect of *Annona senegalensis* on the testicular functions are limited, hence this present study was to evaluate the effect of aqueous leaf extract of *Annona senegalensis* on selected testicular function indices.

## 2. Materials and Methods

### 2.1. Chemicals and Reagents

The following substances were employed: Epinephrine, and hydrogen peroxide ( $H_2O_2$ ), (Sigma® Chemical Company, London, UK); Assay kits for Total Cholesterol (RANDOX® Laboratories Ltd., Antrim, UK). All other reagents used in the study were of analytical grade and highest purity.

### 2.2. Plant Material and Authentication

*Annona senegalensis* leaves were collected from Landmark University farm at Omu-aran. They were identified and authenticated at the Herbarium of the Department of Plant biology, Faculty of Life Science, University of Ilorin. The plant was assigned a voucher number UIH002/1084.

### 2.3. Preparation of the Aqueous Leaf Extract of *Annona senegalensis* (ALEAS)

*Annona senegalensis* leaves were washed with clean water and air dried at room temperature. The leaves were milled into powder; 500 g of the powder was macerated in one litre of distilled water for 24 hours. The mixture was filtered and the filtrate were concentrated using rotary evaporator at 60°C. The concentrated filtrate were collected in a bottle and kept at room temperature before used. The concentrate was reconstituted in distilled water to prepare the various doses of ALEAS used in the study.

### 2.4. Animal Selection and Care

Twenty male Wistar rats weighing between 125 and 127 g were procured from the Experimental Animal Unit of the Department of Biological Sciences, Landmark University, Omu-aran Nigeria. They were acclimatized under laboratory conditions prior to the commencement of experiment. The animals were housed in wire-meshed cages and provided with food and water ad libitum. The animals were kept at standard conditions of temperature and humidity with 12 h light/dark cycles. They were fed with commercial rat diet. Handling of the experimental animals also conforms to international guidelines on the care and use of laboratory animals (National Research Council) [16].

### 2.5. Animal Grouping

The animals were randomly grouped into A, B, C, and D. Animals in group A served as control and were administered 0.5 mL distilled water, while animals in groups group B, C, and D were administered 100, 200 and 300 mg/ kg body weight (bw) respectively of the aqueous leaf extract of *Annona senegalensis* (ALEAS) by oral gavage. The respective doses of ALEAS were delivered in 0.5 mL solution once daily for a period of 21 days.

### 2.6. Animal Sacrifice and Preparation of Testicular Homogenates

The animals were sacrificed under mild diethyl ether anaesthetization 24 hours after cessation of last treatment. The testes were removed and dropped into an isotonic solution, the testes were weighed and homogenized in ice cold 0.25 M sucrose solution (1:5 w/v). The homogenates were centrifuged at 1500 ×g for 10 minutes in a refrigerated centrifuge TDL-5000B, centrifuge (Shanghai Anke company, Ltd., China). The supernatant was collected and stored frozen.

### 2.7. Estimation of Biochemical Indices

Total protein in testicular homogenate was determined by the method of Gornall *et al.* [17]. Catalase (CAT) activity was determined according to the method described by Sinha [18]. The superoxide dismutase (SOD) activity was determined by the method described by Misra and Fridovich [19]. Thiobarbituric acid reactive substances (TBARS) was

measured as an estimate of malondialdehyde (MDA) which is a product of lipid peroxidation using the method described by Satoh [20]. Glycogen content in the testes was determined by the method described by Kemp *et al.* [21]. The concentration of total cholesterol in the testes was analysed using reagent assay kit according to the method of Fredrickson *et al.* [22]. Both acid phosphatase (ACP) and alkaline phosphatase activities (ALP) were determined by the method described by Wright *et al.* [23, 24].

### 2.8. Histological Examination

The preparation of tissues for histological examination was done as described by Adeyemi and Akanji [25]. The representative portions of the testes removed from sacrificed rats were fixed in 10% buffered formalin (pH 7.4) for 12 hrs, then embedded in paraffin. The paraffin embedded tissues were cut into 5-µm sections. The tissue sections were deparaffinised and stained with haematoxylin and eosin. The stained sections were viewed under light microscope and were captured using Bresser DSC-W35 (Meade instruments, Berlin, Germany)

### 2.9. Data Analysis

Data were analysed using one way ANOVA, followed by Duncan post hoc mean comparison test which was used to assess for significant differences between variables. The analysed data were presented as mean of five replicates ± standard error of mean (SEM). P-values less than 0.05 ( $P < 0.05$ ) were considered statistically significant. All statistical analyses were done using the Statistical Package for Social Science (IBM SPSS Statistics 19).

## 3. Results

### 3.1. Phytochemical Constituents of ALEAS

The phytochemical analysis carried out on ALEAS indicated the presence of steroids, terpenoids, anthraquinone, tannins and flavonoids.

### 3.2. Effect of ALEAS on Testis – Body Weight Ratio and Testicular Total Protein in Rat

There was a significant decrease ( $P < 0.05$ ) in the testes-body ratio in the groups treated with ALEAS in a dose dependent fashion when compared to the control (Fig. 1). Similarly there was a decrease in the total testicular protein concentration in group administered various doses of but only found significant in the group administered 300 mg /kg of the ALEAS (Fig 2).

### 3.3. Effect of ALEAS on Testicular Cholesterol, Glycogen and MDA Content in Rat

There was a significant increase ( $P < 0.05$ ) in the testicular total cholesterol (Fig.3) and testicular glycogen (Fig 4) in all the test groups compared to control group. Administration of ALEAS also caused a significant increase in testicular

malondialdehyde concentration in a dose dependent manner (Fig. 5).

### 3.4. Effect of ALEAS on Testicular SOD and CAT Activities in Rat

Activities of testicular enzymic antioxidants, SOD and CAT were increased in a dose – dependent manner following the administration of various doses of ALEAS compared to the control (Fig 6 and 7).

### 3.5. Effect of ALEAS on Testicular ALP and ACP Activities in Rat

Fig. 8 and Fig. 9 represent the influence of different doses of ALEAS on testicular ALP and ACP activities in rat. An increase in both ALP and ACP activities were observed in the ALEAS – treated groups compared to control, although, not significantly different from control ( $P < 0.05$ )

### 3.6. Effect of ALEAS on Testicular Histology in Rat

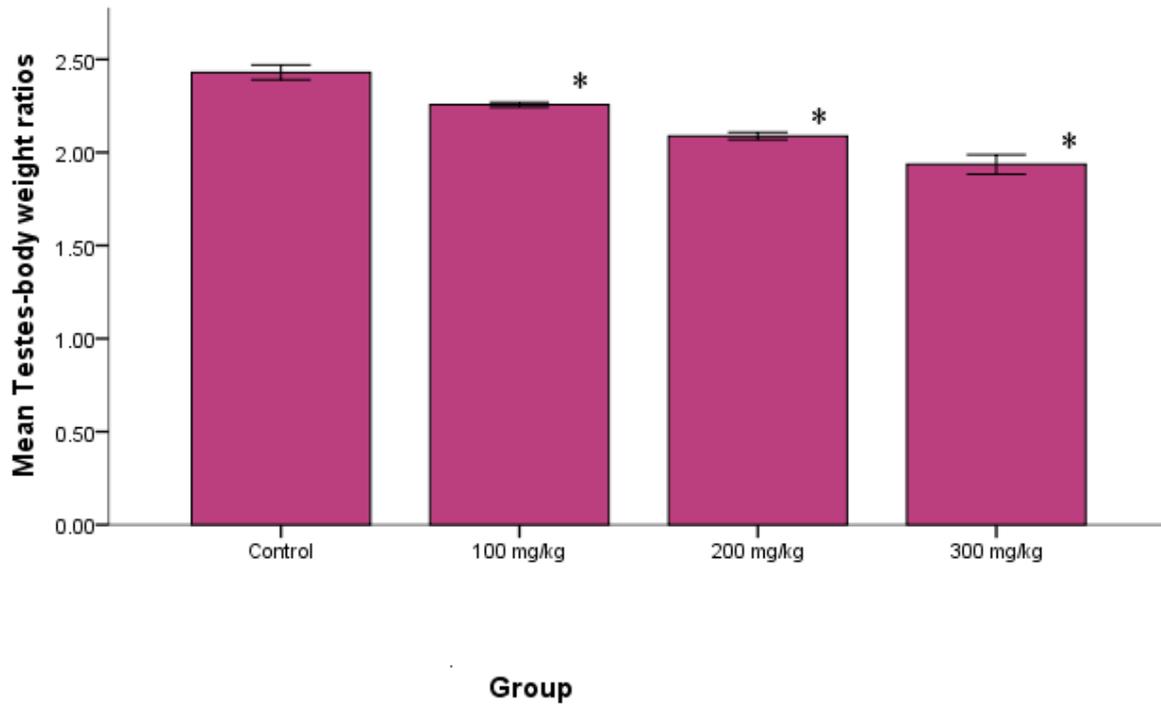
Histopathological data showed a marked degeneration of the seminiferous tubules (Fig. 10). The tissue degeneration were dose dependent with more effect in the group treated with 300 mg/ kg bw ALEAS.

## 4. Discussion

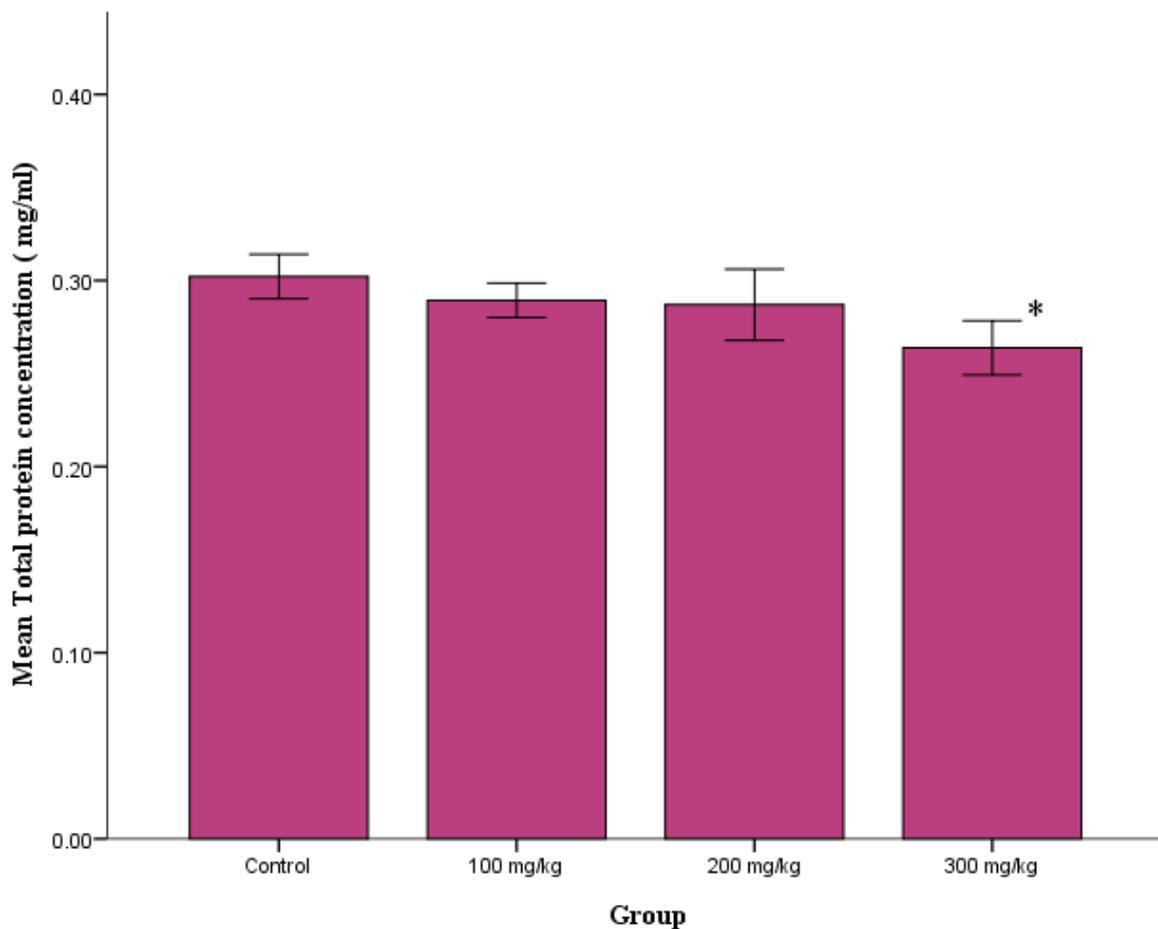
The various parameters investigated in this study are useful indices of evaluating testicular function in experimental animals [26]. A reduction in testis-body weight ratio (Fig.1) observed in the rats administered with the leaf extract suggests the degenerating capacity of ALEAS and this reduction may be due to inhibition of seminiferous tubule fluid formation and loss of germ cells by direct inhibition on spermatogenesis. Our observation regarding reduction in testis - body weight ratio agrees with earlier report on rats [27] and mice [28]. Testicular secretory constituents like protein, cholesterol, sialic acid and glycogen can be used to determine the functional capacity of the testes [29, 30, 31]. Cholesterol is normally secreted into the seminal plasma by the prostate [32] and its function is to protect the spermatozoa against environmental shock. The increase in testicular cholesterol (Fig. 3) in this experiment could be as a result of the alteration in the distribution of lipids in the gonad and increased mobilization from the membrane of testicular cells or increased prostatic secretion of cholesterol into the seminal plasma [33]. The increase in the cholesterol could also result from phagocytosis of sertoli cells hence leaving behind the lipid of the cell membrane [34]. High level of testicular cholesterol has been linked to inhibition of androgen synthesis by leydig cell. Leydig cells release a class of hormones called androgens: testosterone, androstenedione and dehydroepiandrosterone (DHEA), when stimulated by pituitary luteinizing hormone (LH). LH increases the activity

of the enzyme cholesterol desmolase stimulating synthesis and secretion of testosterone by the Leydig cells [35].

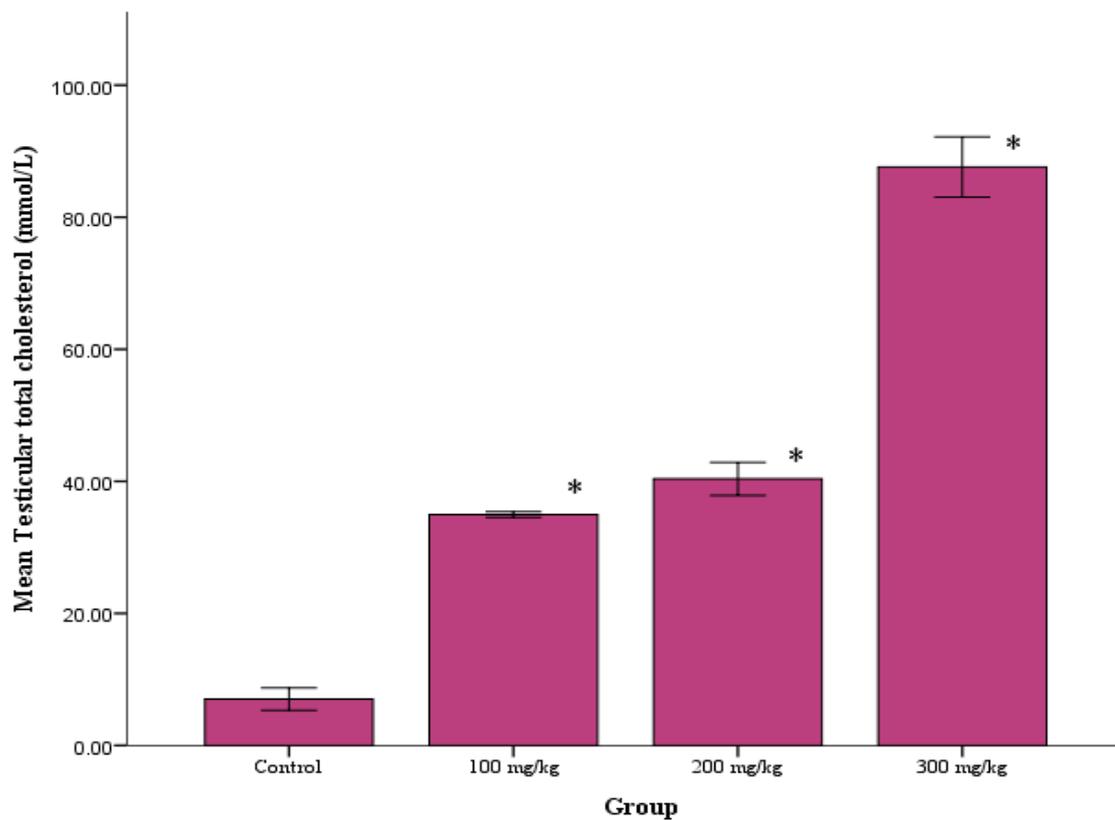
The increase in glycogen (Fig.4) which is the testicular source of glucose could be as a result of interruption in the glucose metabolism or of glucose transport which is the main source of energy for the synthesis of protein in the germinal cells. Glucose transporters are the main transport pathway of glucose to the seminiferous tubules (STs) and spermatogonia [36]. The degeneration caused by the ALEAS could have resulted in the interruption of glucose metabolism and ultimately lipid accumulation in the cytoplasm of the cells especially those located close to lumen of the seminiferous tubules. Malondialdehyde is a metabolic product of a lipid peroxidation. Excessive lipid peroxidation destroys the structure of lipid matrix in the membranes of spermatozoa, and may lead to loss of sperm cell motility [37]. The marked increase in the concentration of testicular MDA (Fig.5) may be due to promotion of peroxidation of testicular membrane lipids by the extract. The antioxidant enzymes such as SOD and CAT are always mobilized against free radicals that may cause lipid peroxidation. SOD is present various cell types where it scavenges superoxide radicals and protects cells from oxidative damage. It has been shown that the activity of antioxidant enzymes behaves in two different ways during oxidative stress. At the beginning of stress, this activity increases, while in the long term, it is reduced due to the massive production of free radicals. This reduction is the result of damage to the molecular principles that is required to induce these enzymes [38, 39]. The non - significant increase in the SOD activity (Fig. 6) could be due to progressive damage to the molecular machinery that is require to induce this enzyme. CAT is efficient antioxidant enzyme and it reduces H<sub>2</sub>O<sub>2</sub> formed in cells to water and molecular oxygen, and it is not saturated by H<sub>2</sub>O<sub>2</sub> at any concentration [40]. The significant increase in catalase activity (Fig. 7) could be in response to an increase in reactive oxygen species due to oxidative stress induced by ALEAS. ALP is involved in mobilizing carbohydrates and lipid metabolites to be utilized either within the cells of the accessory sex structures or by the spermatozoa in the seminal fluid [41]. ACP is widely distributed in the testes and is important in the physiology of sperm [42]. Administration of the graded doses of ALEAS did not show any significant increase in the activities of ALP and ACP (Fig 8 and 9) which may imply that the functions of the enzyme were not significantly altered by the extract during the duration of administration. The histopathological result (Fig 10) showed tissue degenerating effect that was in a dose dependent manner with the group with highest doses showing more degree of degeneration. The extract might have induced apoptosis which is consistent with the histological changes and tissue degeneration [43]. The photomicrograph also revealed a marked depletion of seminiferous tubules and other testicular cells.



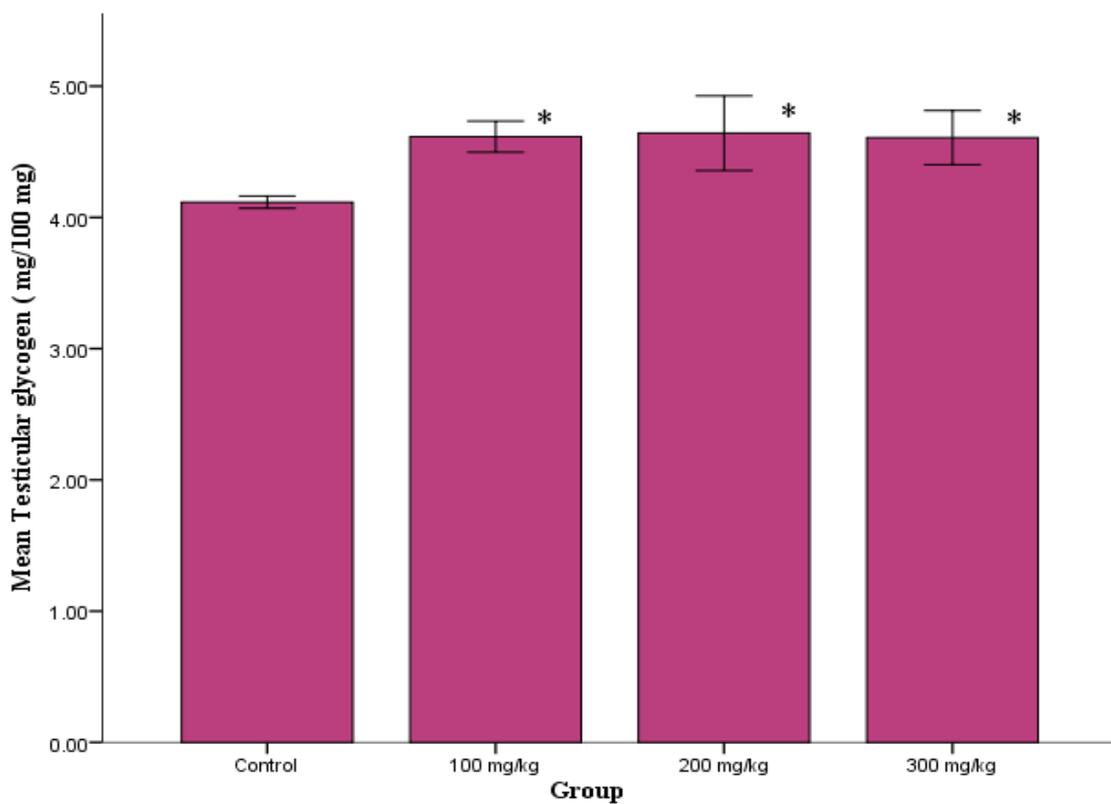
**Fig. 1.** Effect of varying doses of aqueous leaf extract of *Annona senegalensis* on the testes-body weight ratio of the rats. Each value is represented as mean of five replicates  $\pm$  SEM. \*Statistical difference relative to control at  $P < 0.05$ .



**Fig. 2.** Effect of varying doses of aqueous leaf extract of *Annona senegalensis* on the testicular total protein concentration in the rats. Each value is represented as mean of five replicates  $\pm$  SEM. \*statistical difference relative to control at  $P < 0.05$ .



**Fig. 3.** Effect of varying doses of aqueous leaf extract of *Annona senegalensis* on the testicular total cholesterol in the rats. Each value is represented as mean of five replicates  $\pm$  SEM. \*statistical difference relative to control at  $P < 0.05$ .



**Fig. 4.** Effect of varying doses of aqueous leaf extract of *Annona senegalensis* on the testicular total glycogen in the rats. Each value is represented as mean of five replicates  $\pm$  SEM. \*statistical difference relative to control at  $P < 0.05$ .

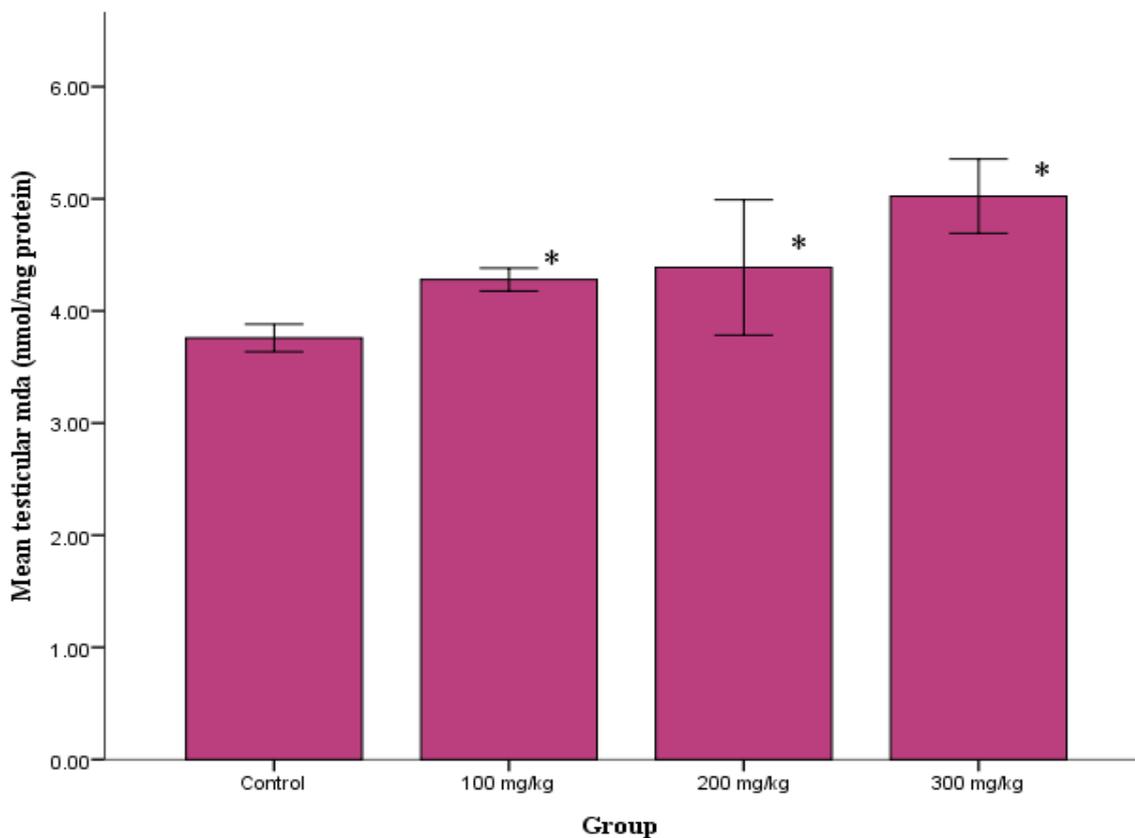


Fig. 5. Effect of varying doses of aqueous leaf extract of *Annona senegalensis* on the testicular malondialdehyde in the rats. Each value is represented as mean of five replicates  $\pm$  SEM. \*statistical difference relative to control at  $P < 0.05$ .

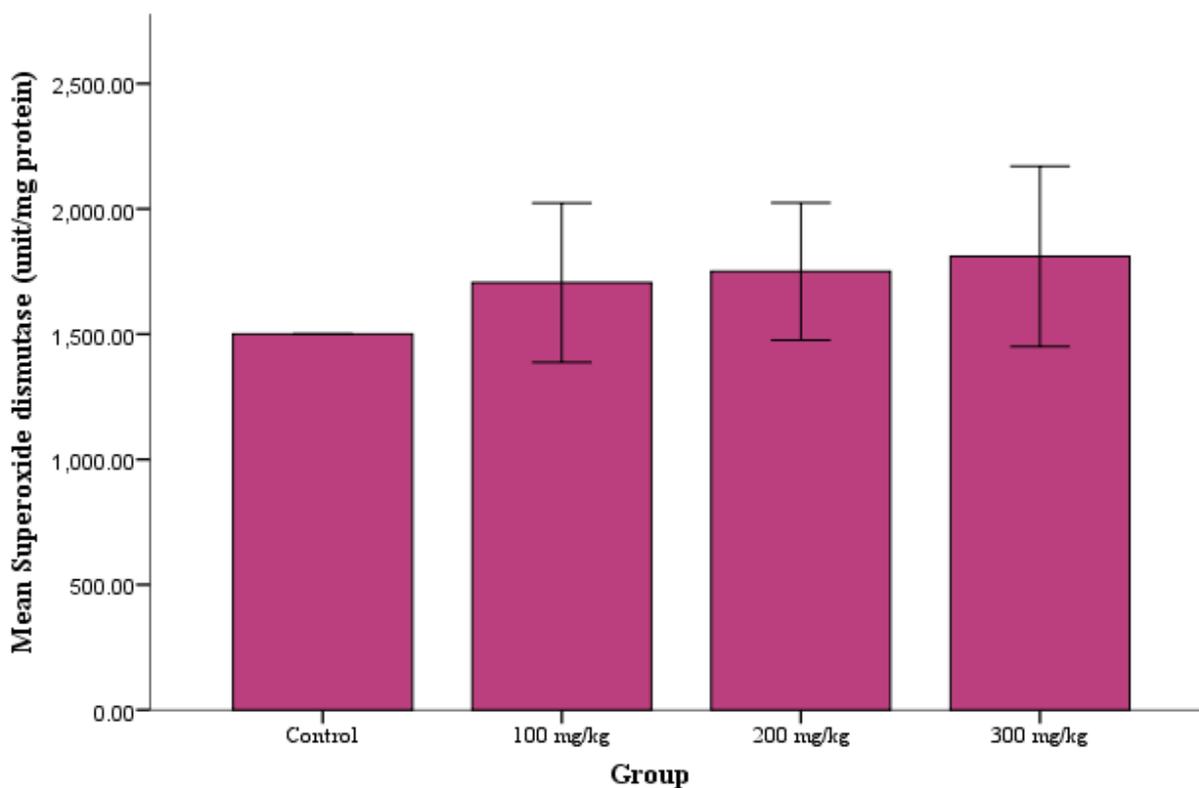


Fig. 6. Effect of varying doses of aqueous leaf extract of *Annona senegalensis* on the testicular malondialdehyde in the rats. Each value is represented as mean of five replicates  $\pm$  SEM. \*statistical difference relative to control at  $P < 0.05$ .

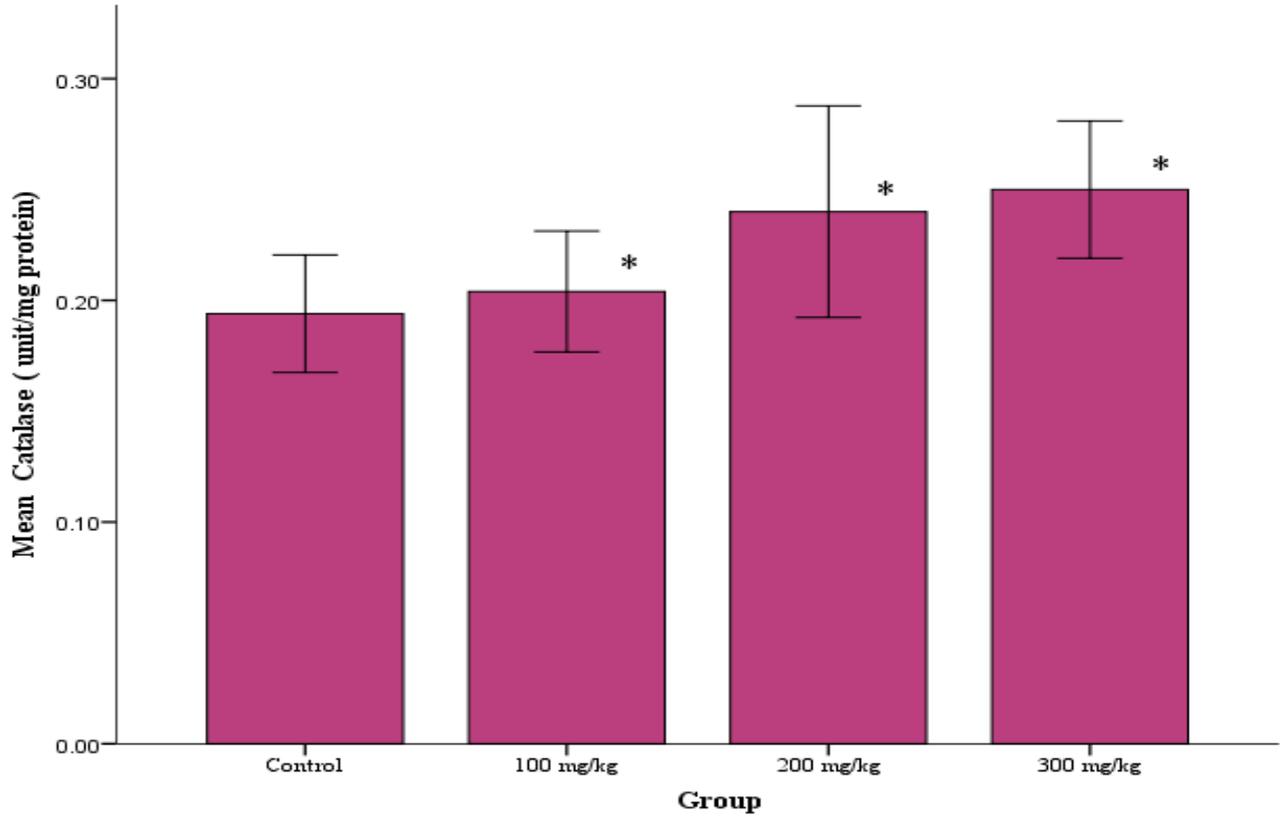


Fig. 7. Effect of varying doses of aqueous leaf extract of *Annona senegalensis* on superoxide dismutase activity in the rats testes. Each value is represented as mean of five replicates  $\pm$  SEM. \*statistical difference relative to control at  $P < 0.05$ .

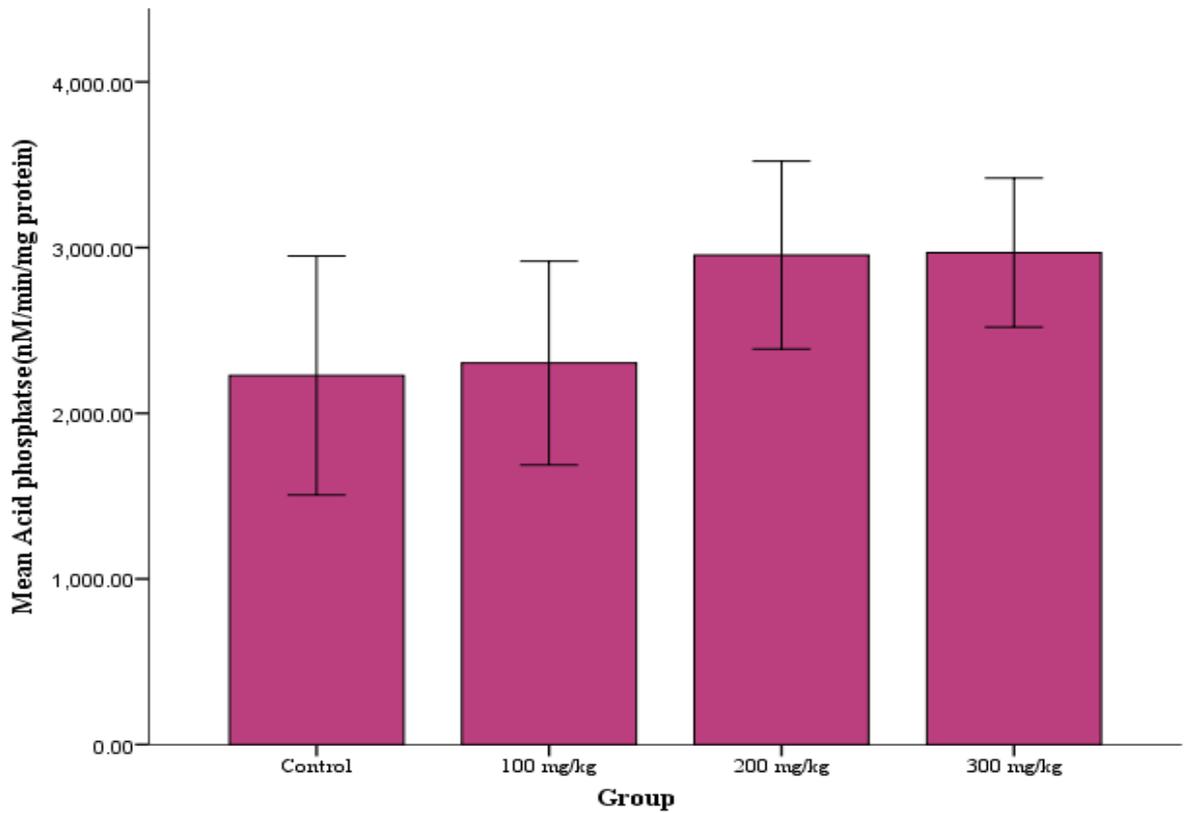
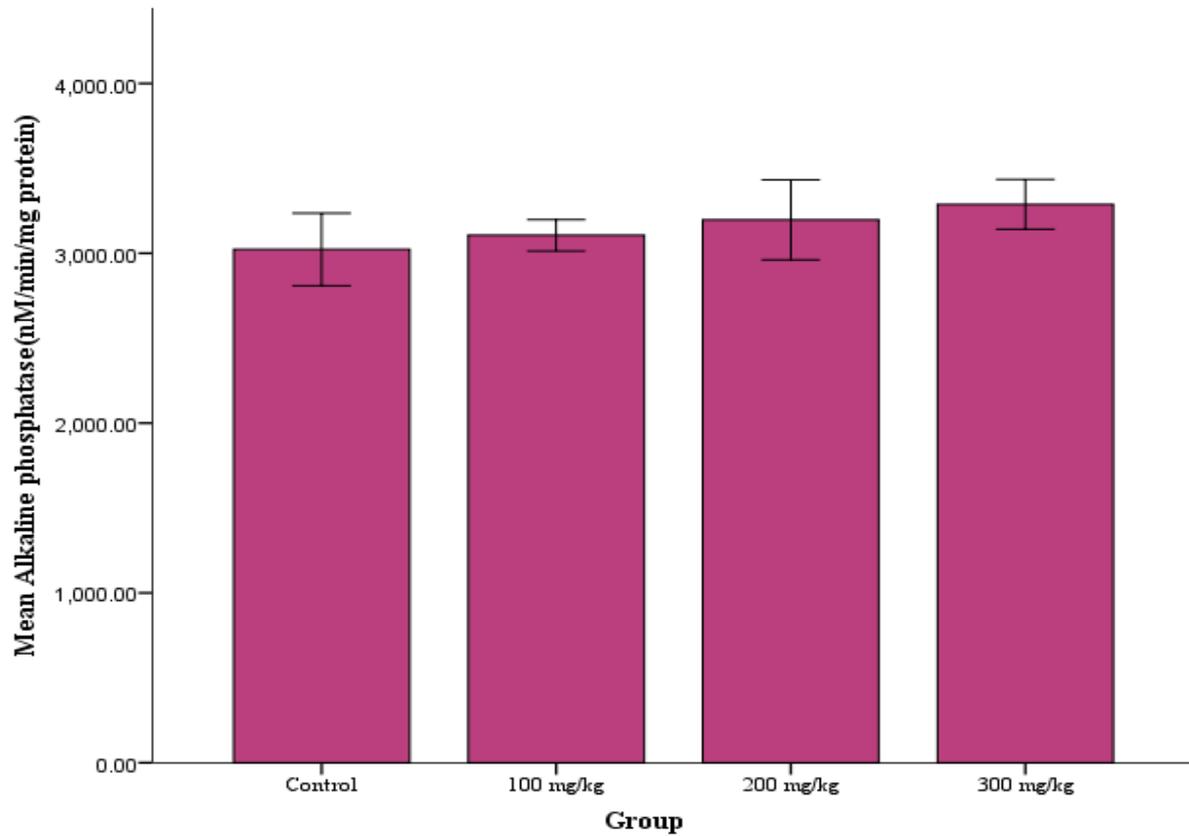
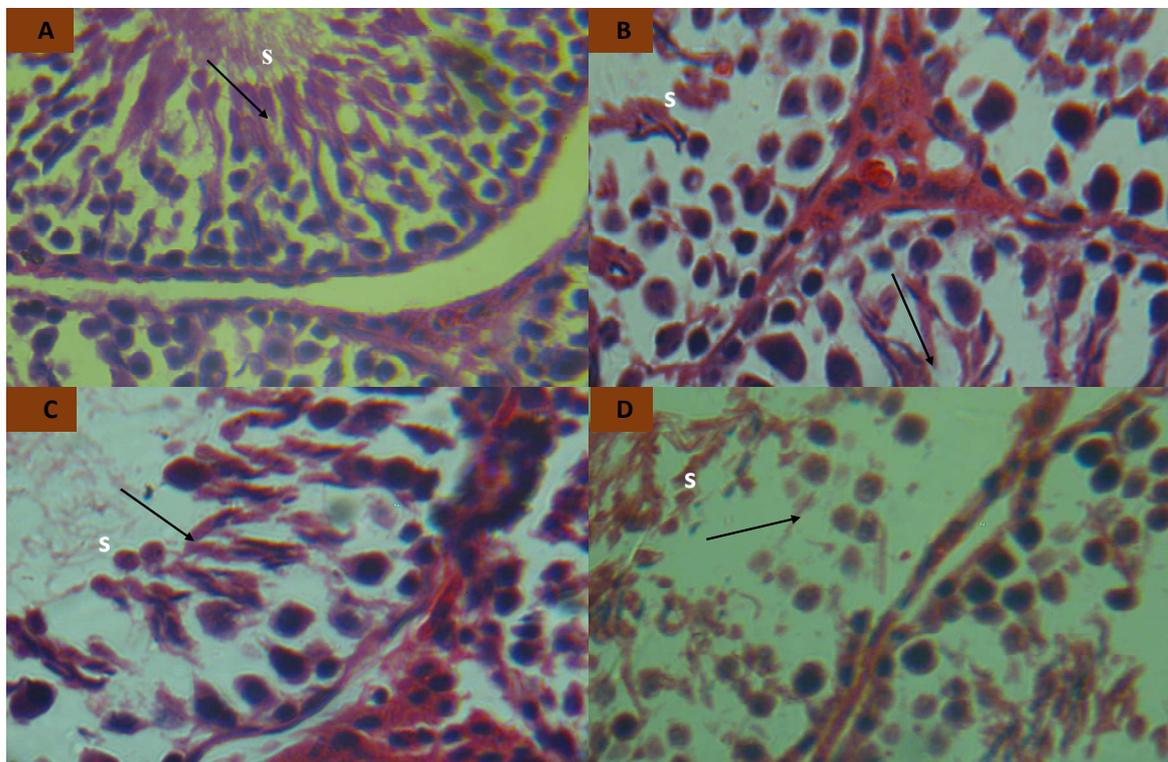


Fig. 8. Effect of varying doses of aqueous leaf extract of *Annona senegalensis* on Acid phosphatase activity in the rats testes. Each value is represented as mean of five replicates  $\pm$  SEM. \*statistical difference relative to control at  $P < 0.05$ .



**Fig. 9.** Effect of varying doses of aqueous leaf extract of *Annona senegalensis* on Alkaline phosphatase activity in the rats testes. Each value is represented as mean of five replicates  $\pm$  SEM. \*statistical difference relative to control at  $P < 0.05$ .



**Fig. 10.** Photomicrographs ( $\times 400$ , H & E) of rat testes (A) Group administered with distilled water (Control); showed testicular tissue with active seminiferous tubules containing cells at various stages of development (spermatogenic series). The cells are normochromic and normocytic. (B) Group administered with 100mg/kg; showed seminiferous tubules with wider lumen. (C) administered with 200mg/kg; showed mildly depleted seminiferous tubules dilated and congested vascular channels. (D) Group administered with 300mg/kg; showed completely depleted seminiferous tubules.

## 5. Conclusion

In the present study, we have established that aqueous leaf extract of *Annona senegalensis* is capable of causing alterations in some biochemical and testicular function parameters considered. These alterations are mostly dose dependent, and are likely indications that the extract may not be completely safe for oral medications.

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